

Robust Inactivation of Yellow Fever Virus 17D Vaccine Strain can be achieved by Photochemical Treatment of Platelet Concentrates

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Contents

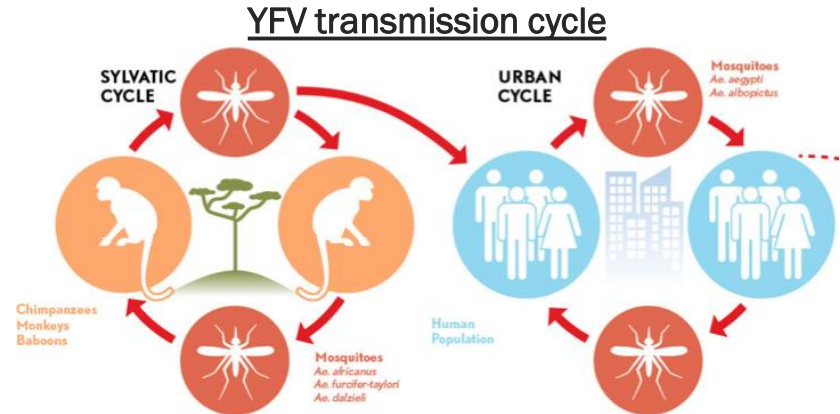
- Yellow Fever Virus (YFV) characteristics, distribution and outbreaks
- Yellow Fever: a threat to the blood supply
- The INTERCEPT™ Blood System
- Pilot YFV inactivation studies
- Inactivation of arboviruses

Disclaimer:

I am an employee of Cerus. Cerus is commercializing the INTERCEPT™ Blood System.

Yellow Fever Virus (YFV)

- Virology:
 - *Flavivirus* / *Flaviviridae*
 - Like Dengue, West Nile, Zika virus
- Clinical outcomes:
 - Mild
 - Hemorrhagic fever
 - Neurotropic / Viscerotropic
- Transmission:
 - Sylvatic (mosquitoes / monkeys)
 - *Haemagogus* sp, *Sabethes* sp,
 - Urban (mosquitoes / humans)
 - *Aedes* sp (*aegypti* and *albopictus*)
 - Transfusion-transmission



YFV mitigation strategies

- Epidemiology:
 - Sporadic in South America / Africa
 - Sylvatic transmission (~300 cases/year)
 - Travelers returning from endemic areas
 - Angola 2015-16 / Brazil 2016-18
- No effective antivirals / only supportive
 - Plasma exchange
 - Liver transplantation
- Vaccine (difficult to produce - limited world supply)
 - Live attenuated 17D-YFV vaccine strain
 - Effective
 - Adverse events (Neurologic / visceral invasion)
 - Risk for immunocompromised patients

National Immunization Schedule

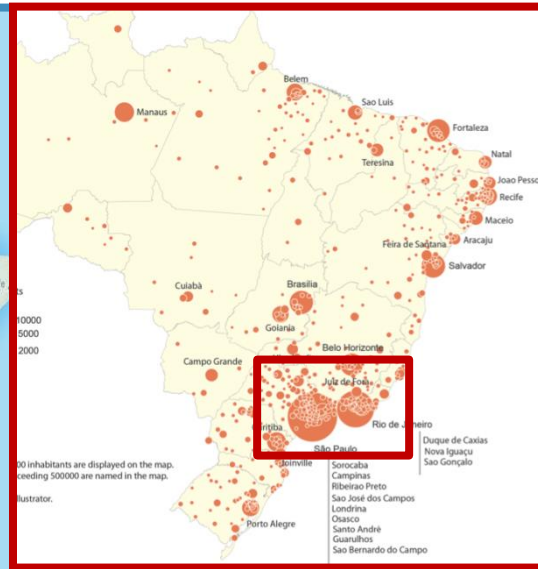
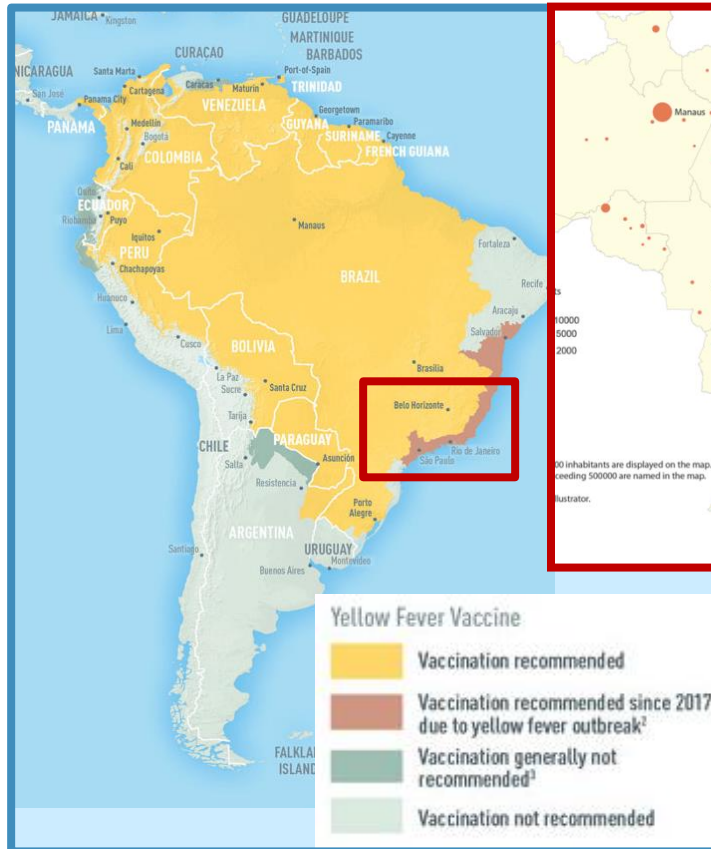


11 | Immunization schedule 2011

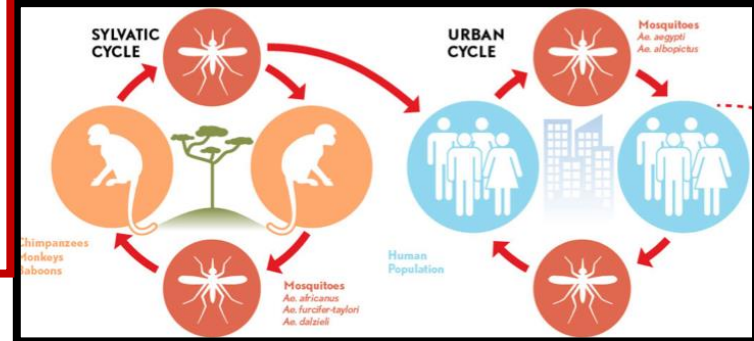


LATAM at risk areas

2016 - 2018 Brazil YFV outbreak



- June 2016 – May 2018:
- YFV was responsible for
 - 2,034 cases
 - 655 fatalities (32%)
 - Many more unrecognized



- A massive vaccination campaign was undertaken (36.7 million doses delivered)
 - To control the outbreak

First transfusion-transmitted YFV infections in Brazil

| Age | Sex | Previous YF vaccine (year) | Blood product received (quantity) | Underlying medical conditions | Symptoms and laboratory abnormalities | YFV IgM | No. of days post-transfusion |
|---------|-----|----------------------------|-----------------------------------|-------------------------------|---------------------------------------|----------------|------------------------------|
| Neonate | F | No | Irradiated RBC (4 aliquots) | Prematurity, IVH | None | Neg | 37 |
| 6 yrs | M | No | Irradiated PC (1 unit) | Wilm´s tumor | None | Pos/ 160 | 36 |
| 66 yrs | M | Yes (1964) | PC (1 unit) | Kidney transplant, diabetes | None | Pos/ 160 | 33 |
| 58 yrs | M | Yes (1975, 1986) | FFP (2 units) | CRF, tuberculosis, psoriasis | None | Pos/ 40,960 | 26 |
| 82 yrs | M | Yes (1959, 1965) | Irradiated PC (1 unit) | B-cell lymphoma | Deceased | NA | - |

Risk of transfusion-transmitted YFV vaccine

- MMWR Jan. 22, 2010 - Evidence of YFV vaccine TTI in the US military
 - 89 military personnel donated blood 4 days after vaccination
 - 6 units were transfused to 5 patients: 1 died, others seroconverted
- Live attenuated virus → viremia after vaccination

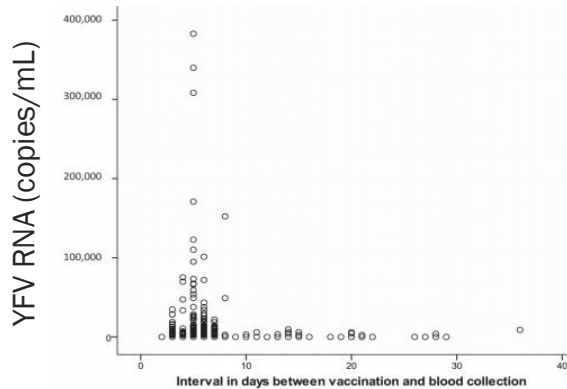


Table 7. Vaccine virus RNA positivity by RT-PCR at specific intervals between vaccination and blood sampling

| | < 11 d | ≥ 11 d to < 14 d | ≥ 14 d |
|-------------------|--------|------------------|--------|
| N | 787 | 19 | 65 |
| % RT-PCR positive | 42.3% | 10.5% | 16.9% |

Viremia up to 36 days after vaccination

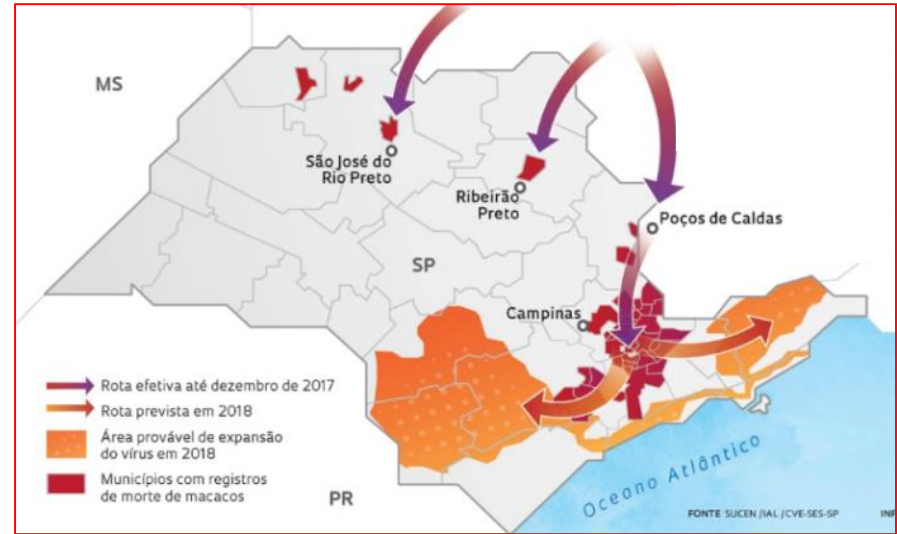
Martins RMM. Human Vaccines & Immunotherapeutics 2013

Data presented by Dr Luiz Amorim, Hemorio at SVTM Swiss congress 2017

- Blood donor deferral period of 2-4 weeks to limit the risk of YFV vaccine TTI

YFV: a threat to the blood supply

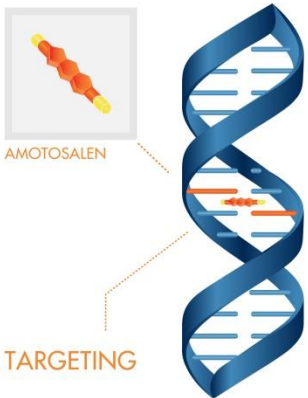
- YFV TTI documented for:
 - Wild type strain
 - Live attenuated vaccine strain
- Deferral based on clinical signs:
 - 80% asymptomatic cases
 - Difficult to prevent donation from asymptomatic donors
- Deferral based on risk after vaccine
 - Immunocompromised patients at higher risk for severe outcome



- Deferral policies may increase the risk of platelet shortage in areas impacted by massive vaccination campaign
- **Pathogen inactivation could be an alternative strategy to help mitigate the risk of YFV TTI and platelet shortage**

INTERCEPT Mechanism of Action Targeting DNA and RNA to Prevent Pathogen Proliferation

1 Intercalates Into Regions of DNA and RNA



2 Crosslinks Upon UVA Illumination



3 Blocks Replication, Transcription and Translation



- 1) Amotosalen, a psoralen, penetrates cellular and nuclear membranes and intercalates into helical regions of DNA or RNA.
- 2) Amotosalen forms covalent crosslinks to nucleic acid base pairs upon exposure to UVA light.
- 3) DNA and RNA replication is blocked. Pathogens and leukocytes cannot replicate and are 'inactivated.'

The INTERCEPT Blood System for Platelets

INTEGRATED CONTAINER SET



STEP 1
Amotosalen

STEP 2
Illumination

STEP 3
CAD

Process Complete
Storage



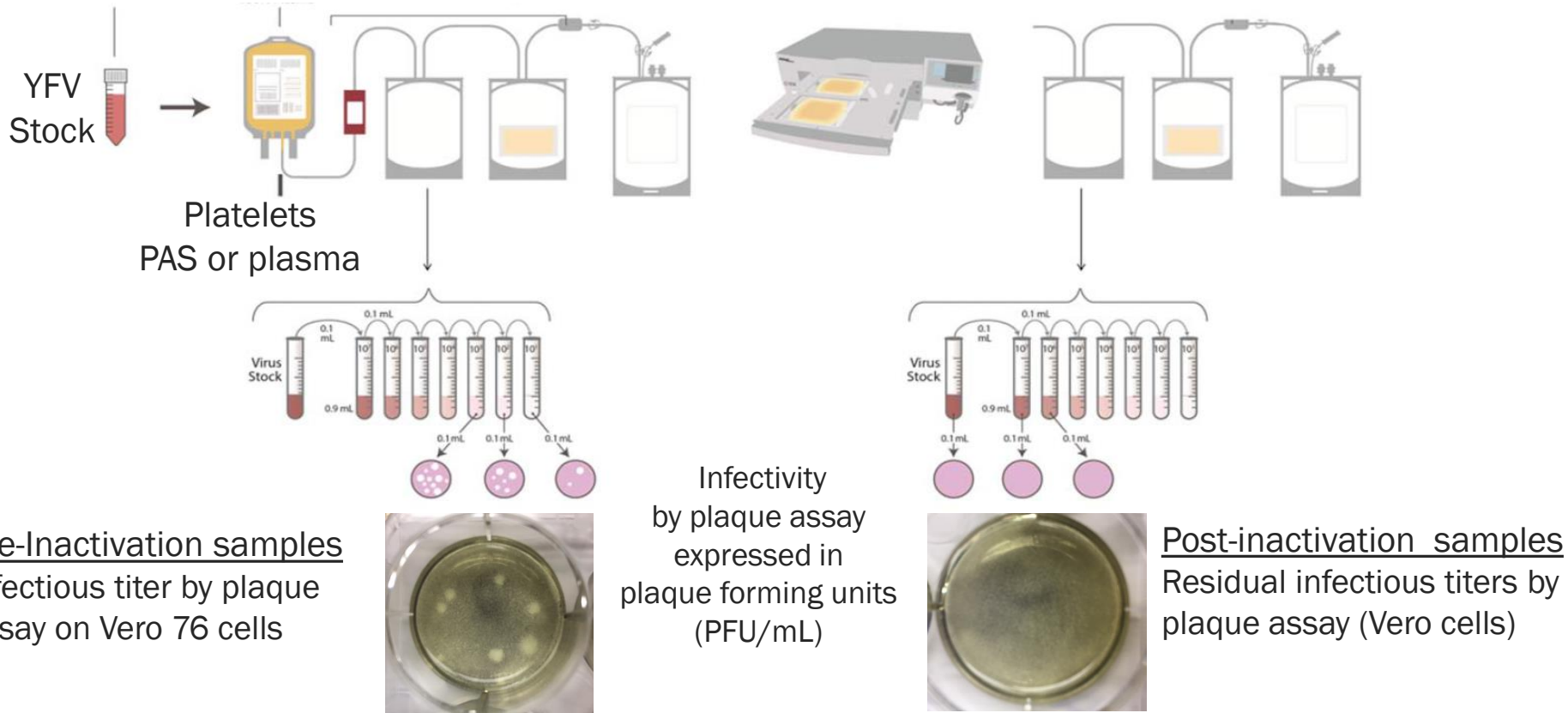
**UVA
Illumination
Device**

- Using a sterile connecting device (SCD), the platelet container is connected to the INTERCEPT kit.
- Amotosalen (1) is added by gravity flow and the platelet mixture is illuminated with UVA light (2).
- Residual amotosalen and its photoproducts in the platelet mixture are reduced to low levels using a compound adsorption device (CAD) (3) before the platelets are transferred to the storage containers.

Pilot study objectives

- Evaluate the ability to inactivate 17D-YFV using amotosalen (S-59) and UVA light for pathogen inactivation treatment of platelet components (PC)
- Use infectivity assays with Vero76 cells

YFV vaccine inactivation – Experimental design



YFV vaccine inactivation - Results

| | PC in 35% plasma / 65% PAS (n=4*) | PC in 100% plasma (n=1*) |
|---|--------------------------------------|-----------------------------|
| Pre-PI Treatment (Log ₁₀ PFU/mL) | 4.65 ± 0.6 | 5.19 |
| Post-PI Treatment (Log ₁₀ PFU/mL) | <-0.70 ± 0.0 | <-0.7 |
| Log reduction (Log ₁₀ PFU/mL) | >4.65 ± 0.6 | >5.19 |
| Log reduction (Log ₁₀) | >5.34 ± 0.6 | >5.89 |

*n=4 more replicates in PAS and 100% plasma are planned for regulatory submission

Inactivation of Arboviruses with the INTERCEPT Blood Systems to the Limit of Detection

| Arbovirus | Log ₁₀ Inactivation | | |
|---------------------|--------------------------------|--------------------|--------------------|
| | Plasma | Platelets | RBC* |
| WNV | ≥6.8 ⁴ | ≥6.0 ⁵ | NA |
| DENV | >5.6 ¹ | ≥4.3 ⁶ | >6.6 ¹⁰ |
| CHIKV | ≥7.6 ⁷ | >6.4 ⁷ | > 7.1 ⁸ |
| ZIKV** | >6.6 ² | >4.9 ⁹ | >5.8 ³ |
| °YFV (strain 17D)** | NA | >5.9 ¹¹ | NA |
| °MAYV** | NA | >6.9 ¹² | >6.2 ¹² |
| °RRV** | NA | >5.1 ¹³ | >5.5 ¹³ |

1. Musso et al. Transfusion 2014;54: 2924-30.

2. Aubry et al. Transfusion 2016;56: 33-40.

3. Laughhunn et al. Transfusion 2017 Feb 5. doi: 10.1111/trf.13993.

4. Lin L, Hanson CV, et al. Transfusion 2005;45(4): 580-590.

5. Singh Y, Sawyer LS, et al. Transfusion 2006;46(7): 1168-1177.

6. Dupuis K, Arnold DJ, Sawyer L. Transfusion 2012;52(3S):225A.

7. Tsetsarkin et al. Am J Trop Med Hyg 2013;88: 1163-9.

8. Laughhunn et al. Transfusion 2017;58:748-757.

9. Santa Maria et al., Transfusion 2017 Aug;57(8):2016-2025.

10. Aubry, et al., TRANSFUSION 2017;57:2888-2896.

11. Laughhunn, et al., Robust Inactivation of the Yellow Fever Virus 17D Strain Can Be Achieved Using Amotosalen and UVA Light for Pathogen Inactivation Treatment (PRT) Of Platelet Components – AABB 2017.

12. Santa Maria, et al., Robust Inactivation of Mayaro Virus in Platelet Concentrates and Red Blood Cells using nucleic acid targeting pathogen reduction technologies (PRT) – AABB 2017.

13. Laughhunn, et al., Effective inactivation of Ross River virus in blood components through nucleic acid targeting – AABB 2017.

* INTERCEPT Blood System for RBC is in development and not approved for commercial use.

**Data for pathogen inactivation of Zika, Yellow Fever, Mayaro, or Ross River Viruses by INTERCEPT Blood System have not been submitted to TUV or FDA for review.

Arbovirus Comparison

| Characteristic | Dengue ¹ | Chikungunya ² | Zika ^{3,4} | West Nile ⁵ | Yellow Fever ⁶ |
|------------------------------|---|---|--|--|---|
| Virus family | Flaviviridae | Togaviridae | Flaviviridae | Flaviviridae | Flaviviridae |
| Virus genus | Flavivirus | Alphavirus | Flavivirus | Flavivirus | Flavivirus |
| Serotypes | 4 | 1 | 1 | 1 | 1 |
| Vector | <i>Aedes aegypti</i> <i>Aedes albopictus</i> | <i>Aedes aegypti</i> <i>Aedes albopictus</i> | <i>Aedes aegypti</i> <i>Aedes albopictus</i> | <i>Culex Pipiens</i> <i>C. Tarsalis</i> <i>C. quinequefasciaatus</i> | <i>Aedes aegypti</i> <i>Aedes albopictus</i> |
| Symptomatic: Asymptomatic | 25:75 | 75:25 | 20:80 | 20:80 | ~20:80 |
| Illness outcome | Severe dengue – plasma leakage | Rarely severe, Arthralgias | Rarely severe, Guillain-Barré Microcephaly | Neuroinvasive disease Meningitis Encephalitis Polyomyelitis | Rarely severe, but ~15% of patients progress to serious form; hemorrhagic fever, multisystem organ failure |

1. <http://www.cdc.gov/dengue/>
2. <http://www.cdc.gov/chikungunya/>
3. <http://www.cdc.gov/zika/>
4. Zika Virus: Information for Clinicians, June 13, 2016, <http://www.cdc.gov/zika/pdfs/clinicianppt.pdf>
5. <http://www.cdc.gov/westnile/>
6. <https://wwwnc.cdc.gov/travel/yellowbook/2016/infectious-diseases-related-to-travel/yellow-fever>

Conclusions

- S-59/UVA-mediated PRT is efficient at inactivating the 17D-YFV vaccine strain in PC independently of resuspension medium
 - Similar to results obtained with other Flaviviruses, including West Nile, Dengue and Zika virus
- Because of high rates of asymptomatic infections, donor deferral is an inefficient mitigation strategy for most arboviruses
- Blood recipients are often immunocompromised
 - At risk for the development of severe clinical outcomes
- Pathogen reduction is a potential mitigation strategy to prevent TTI's and maintain PC availability in areas
 - affected by large YFV outbreaks
 - with widespread vaccination campaigns